

## A NOVEL ACYLATED PELARGONIDIN 3-SOPHORO- SIDE-5-GLUCOSIDE FROM GREYISH-PURPLE FLOWERS OF THE JAPANESE MORNING GLORY

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**Abstract** — A novel acylated anthocyanin was isolated from greyish-purple flowers of a *dingy* mutant in the Japanese morning glory (*Pharbitis nil* or *Ipomoea nil*) with three known pigments as main pigments. This novel pigment was determined to be pelargonidin 3-*O*-[2-*O*-( $\beta$ -D-glucopyranosyl)-6-*O*-(*trans*-4-*O*-( $\beta$ -D-glucopyranosyl)caffeoyl)- $\beta$ -D-glucopyranoside]-5-*O*- $\beta$ -D-glucopyranoside (**3**), and the other three pigments (**1**, **2**, and **4**) were 3-sophoroside-5-glucosides of pelargonidin and peonidin, and peonidin 3-*O*-[2-*O*- $\beta$ -D-glucopyranosyl-(6-*O*-( $\beta$ -D-glucopyranosyl)caffeoyl)- $\beta$ -D-glucopyranoside]-5-*O*- $\beta$ -D-glucopyranoside. In addition, pelargonidin 3-sophoroside and pelargonidin 3-glucoside were also detected as main pigments in two other mutants, *duskish-1* and *duskish-2*, displaying pale-purple hue flowers.

Recessive mutations of the Japanese morning glory (*Pharbitis nil* or *Ipomoea nil*) conferring dull colors can be classified into four alleles, *dusky*, *duskish-1*, *duskish-2*, and *dingy*.<sup>1</sup> In our continuing efforts to elucidate flower color variation due to the production and accumulation of anthocyanins in the petals of *Pharbitis nil* cultivars bearing various flower colors,<sup>1-9</sup> we have reported the anthocyanin constituents of pigments in the dull color flowers, such as the maroon and slate flowers, of *Pharbitis nil* lines which are

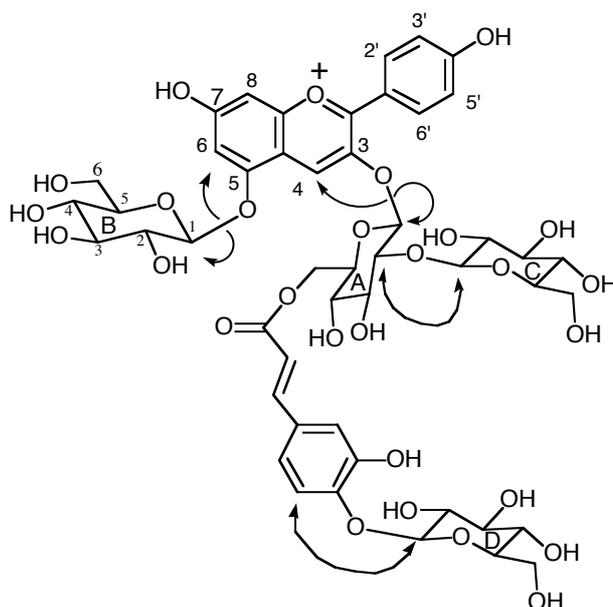
believed to carry the *dusky* mutations.<sup>6,8</sup> The dull color flowers in these apparent *dusky* mutations contained anthocyanin 3-monoglucosides or 3,5-diglucosides as their deacylanthocyanins,<sup>6,8</sup> whereas the corresponding deacylanthocyanins in the bright color flowers of several cultivars examined were anthocyanidin 3-sophoroside-5-glucosides.<sup>3,4</sup> Indeed, the components of anthocyanin pigments in the flowers of an authentic *dusky* mutant<sup>10</sup> were found to be similar to those in the previously characterized cultivar 'Danjuro'.<sup>6</sup>

In this communication, we wish to report the structure of a novel acylated pelargonidin 3-sophoroside-5-glucoside, along with three known anthocyanins, in the grayish-purple flowers of a *dingy* mutant,<sup>1,10</sup> and also the presence of pelargonidin 3-sophoroside and pelargonidin 3-glucoside in the pale-purple hue flowers of the mutants carrying either the *duskyish-1* or *duskyish-2* allele.<sup>1,10</sup>

Dry flowers (15 g) of a *dingy* strain of *Pharbitis nil* were immersed in 5% AcOH for 24 h at room temperature to leave the red extract, in which six anthocyanin peaks were observed as main anthocyanins by HPLC analysis.<sup>11</sup> Their relative frequency of occurrence (the average of 5 HPLC analyses) was pigment-1 (**1**) (18%),<sup>12</sup> -2 (**2**) (7%),<sup>13</sup> -6 (**3**) (58%),<sup>14</sup> and -7 (**4**) (6%).<sup>15</sup> (Figure 1) From these mixed anthocyanins, pigment-6 (**3**) (*ca.* 20 mg), pigment-1 (**1**) (*ca.* 8 mg) and pigment-7 (**4**) (*ca.* 1 mg) were obtained as single compounds, respectively, by the procedure described previously.<sup>9,16,17</sup> The pigment-1 (**1**), -2 (**2**), and -7 (**4**) were elucidated to be pelargonidin 3-sophoroside-5-glucoside, peonidin 3-sophoroside-5-glucoside, and peonidin 3-*O*-[2-*O*-( $\beta$ -D-glucopyranosyl)-6-*O*-( $\beta$ -D-glucopyranosyl)-caffeoyl- $\beta$ -D-glucopyranoside]-5-*O*- $\beta$ -D-glucopyranoside (Pharbitis Blue anthocyanin 2),<sup>4</sup> respectively, by direct comparison of HPLC and TLC with authentic samples.<sup>2-5</sup>

Pigment-6 (**3**) was a novel anthocyanin, which on acid hydrolysis gave pelargonidin, glucose and caffeic acid. The FAB-MS spectra of pigment-6 (**3**) gave its molecular ion [M<sup>+</sup>] at 1081 *m/z*, in good agreement with the mass calculated for C<sub>48</sub>H<sub>57</sub>O<sub>28</sub> (1081.304), which was composed of pelargonidin with four molecules of glucose and one molecule of caffeic acid. This mass value was confirmed by the measurement of its high resolution FAB-MS [Calcd for C<sub>48</sub>H<sub>57</sub>O<sub>28</sub> requires: 1081.3036. Found: 1081.3014].<sup>14</sup> In order to determine the structure of this pigment (Figure 2), its <sup>1</sup>H NMR measurements including <sup>1</sup>H-<sup>1</sup>H COSY and negative NOE difference (DIFNOE)<sup>2</sup> spectra were carried out in CF<sub>3</sub>CO<sub>2</sub>D-DMSO-d<sub>6</sub> (1:9). The signals for ten aromatic protons of pelargonidin and caffeic acid moieties were assigned as shown in Table 1. Two olefinic protons of caffeic acid showed a large coupling constant (*J* = 15.6 Hz) supporting a *trans*

configuration (Figure 2). The proton signals of sugar part (glucose was abbreviated as Glc throughout this paper) were observed in the region of 5.60 - 3.00 ppm (Table 1). The signals of four anomeric protons appeared at 5.66 ppm [d,  $J = 7.2$  Hz, Glc A], 5.11 ppm (d,  $J = 7.2$  Hz, Glc B), 4.75 ppm (d,  $J = 7.6$  Hz, Glc C), and 4.80 ppm (d,  $J = 6.4$  Hz, Glc D), indicating that all the glucose units are  $\beta$ -D-glucopyranosides. The two characteristic protons at 4.34 and 4.42 ppm with a geminal coupling constant ( $J = 11.3$  Hz) were assigned to the methylene (C-6) of Glc A, supporting the notion that the caffeoyl moiety is attached to 6-OH of Glc A. Based on the analysis of its  $^1\text{H}$ - $^1\text{H}$  COSY spectrum, the anomeric proton (5.66 ppm, Glc A) was finally correlated with both protons (4.34 and 4.42 ppm, H-6a and 6b of Glc A). The proton (4.08 ppm, t,  $J = 7.6$  Hz) at the 2-position of Glc A was also shifted in the lower magnetic field, indicating 1-OH of Glc C was bonded with 2-OH of Glc A through a glucosidic bond. This linkage was confirmed by the observation of a strong negative NOE signal at H-2 of Glc A by irradiation of H-1 of Glc C in its DIFNOE spectrum<sup>2</sup> (Figure 2).



**Figure 1** An acylated anthocyanin (Pigments-6) (**3**) from the dingy strain of *Pharbitis nil*. Observed NOE's are indicated by arrows.

Furthermore, the appearance of a negative NOE signal at H-4 of pelargonidin by irradiation at H-1 of Glc A showed Glc A to be attached to 3-OH of pelargonidin through a glycosidic bond, and Glc B was bonded to 5-OH of pelargonidin due to the occurrence of the strong NOEs between H-6 of pelargonidin and H-1 of Glc B. Similarly, Glc D was confirmed to attach to 4-OH of caffeic acid through a glycosidic bond by the presence of NOEs between H-5 of caffeic acid and H-1 of Glc D (Figure 2). Consequently, the structure of pigment-6 (**3**) was unambiguously determined to be pelargonidin 3-*O*-[2-*O*-( $\beta$ -D-glucopyranosyl)-6-*O*-

(*trans*-4-*O*-( $\beta$ -D-glucopyranosyl)caffeoyl)- $\beta$ -D-glucopyranoside]-5-*O*- $\beta$ -D-glucopyranoside, which has not been reported previously in plants.<sup>18,19</sup>

Table 1 <sup>1</sup>H NMR spectral data of a dingy anthocyanin (pigment-6) (**3**) of *Pharbitis nil*.\*

Pelargonidin		Glucose A (3-Glc)		Glucose C (2"-Glc)	
4	8.85	1	5.66 d (7.2)	1	4.75 d (7.6)
6	7.00 br s	2	4.08 t like (7.6)	2	3.00 t like (8.4)
8	7.12 br s	3	3.75 t like (8.8)	3	3.14 t like (8.7)
2'	8.61 d (7.6)	4	3.53 t like (8.2)	4	3.05 t like (9.2)
3'	7.10 d (7.6)	5	3.97 m	5	2.86 m
5'	7.10 d (7.6)	6a	4.34 m	6a	3.28 m
6'	8.61 d (7.6)	6b	4.42 d (11.4)	6b	3.33 m
Caffeic acid		Glucose B (5-Glc)		Glucose D	
2	7.00 br s	1	5.11 d (7.2)	1	4.80 d (6.4)
5	7.11 d (9.2)	2	3.53 m	2	3.33 m
6	6.99 m	3	3.39 m	3	} 3.55 - 3.18
$\alpha$	6.33 d (15.7)	4	3.28 m	4	
$\beta$	7.39 d (15.7)	5	3.51 m	5	
		6a	3.55 m	6a	} 3.75 m
		6b	3.80 d (11.3)	6b	

\* <sup>1</sup>H NMR (500 MHz) (CF<sub>3</sub>CO<sub>2</sub>D-DMSO-d<sub>6</sub>, 1:9), at 25 °C, an internal standard of TMS; Coupling constants (*J* in Hz) in parentheses.

Except for their aglycones, the anthocyanins for acylation and glycosidation in the *dingy* mutant were similar in structure to those in the cultivars with light-blue flowers characterized before.<sup>4</sup> The *dingy* mutant contained pelargonidin and peonidin, while the light-blue cultivars peonidin and cyanidin. It is thus likely that the light-blue cultivars also carry *dingy* mutant(s) and that the *Dingy* gene product is responsible for the acylations of caffeic acids at both 6-OH residues of Glc C and D, since the pigments in both the light-blue cultivars and the *dingy* mutant displaying grayish-purple flowers were lacking the acylations.

In other two pale grayish-purple flower mutants, *duskish-1* and *duskish-2*, the anthocyanin extracts were obtained from their flowers with the same solvents, and their anthocyanin components were analyzed by HPLC with the authentic anthocyanins. As Figure 2 shows, pelargonidin 3-sophoroside (pigment-3)<sup>20</sup> and pelargonidin 3-glucoside (pigment-4)<sup>21</sup> were detected in the extract of *duskish-1* as main anthocyanins. The extract of *duskish-2* contained pelargonidin 3-[tri(glucosylcaffeoyl)sophoroside]-5-glucoside (Pharbitis red anthocyanin 1, pigment-8)<sup>22</sup> and pelargonidin 3-[2-glucosylcaffeoylglucosyl-6-(caffeoylglucosyl-caffeoyl)glucoside]-5-glucoside (Ipomoea red anthocyanin 1, pigment-9)<sup>23</sup> as its main anthocyanins as well as pelargonidin 3-sophoroside and 3-glucoside. Interestingly, total anthocyanin contents of both flowers of the *duskish-1* and *duskish-2* mutants were found to be reduced to less than 10% of that in the flower extract

of the *dingy* mutant. Since the content of anthocyanins in the *dingy* flowers appears to be normal, the results imply that both *duskyish* mutations affect either the biosynthesis of anthocyanin pigments or their transport into the vacuoles of the floral epidermal tissue. Identification of the *Duskyish* gene would help to clarify its role in pigmentation processes and would provide an explanation for the pale pigmentation of the *duskyish* mutants.

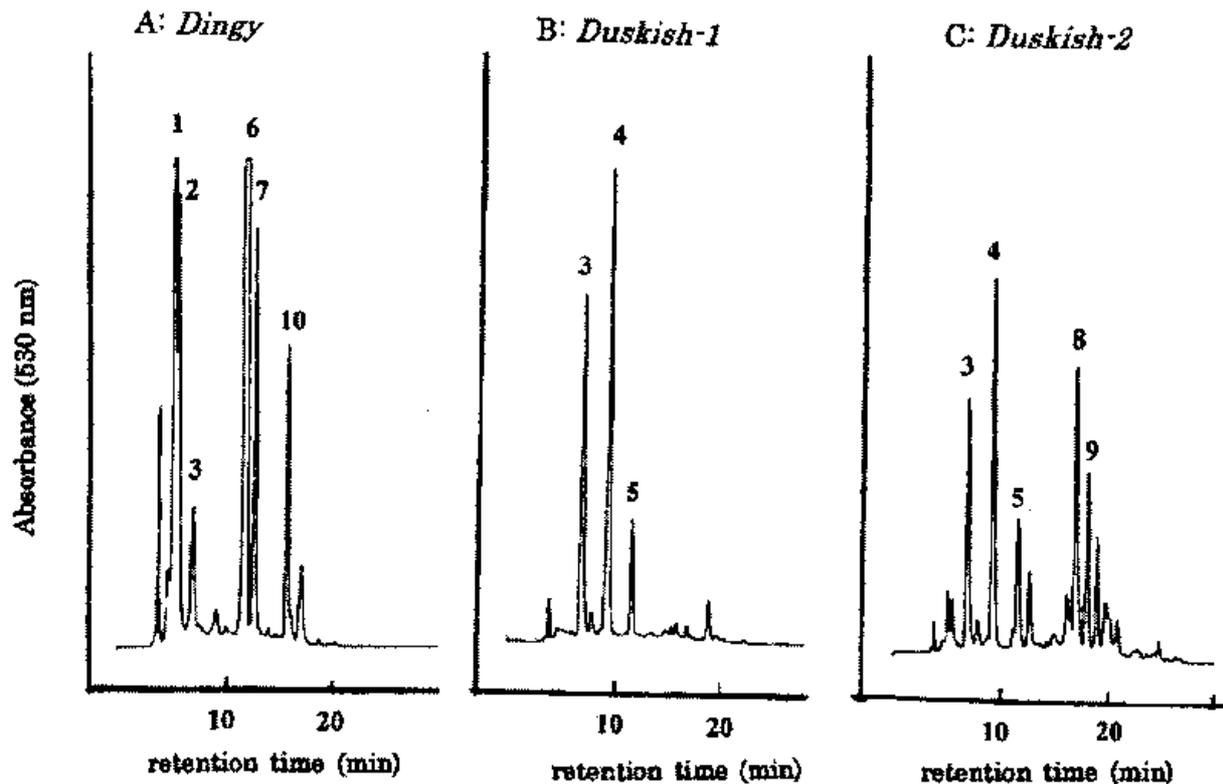


Figure 2. HPLC Analysis of the pigments in the *dingy* (A), *duskyish-1* (B), and *duskyish-2* (C) flowers *Pharbitis nil* mutants. The pigments identified are: pigment-1 (peak 1) = pelargonidin 3-sophoroside-5-glucoside; pigment-2 (peak 2) = peonidin 3-sophoroside-5-glucoside; pigment-3 (peak 3) = pelargonidin 3-sophoroside; pigment-4 (peak 4) = pelargonidin 3-glucoside; pigment-5 (peak 5) = pelargonidin; pigment-6 (peak 6) = pelargonidin 3-[2-glucosyl-6-(4-glucosylcaffeoyl)glucoside]-5-glucoside; pigment-7 (peak 7) = peonidin 3-[2-glucosyl-6-(4-glucosylcaffeoyl)glucoside]-5-glucoside; pigment-8 (peak 8) = pelargonidin 3-[2-glucosylcaffeoylglucosyl-6-(glucosylcaffeoylglucosylcaffeoyl)glucoside]-5-glucoside (*Pharbitis Red* anthocyanin 5); pigment-9 (peak 9) = pelargonidin 3-[2-glucosylcaffeoylglucosyl-6-(caffeoylglucosylcaffeoyl)glucoside]-5-glucoside (*Ipomoea Red* anthocyanin 1); pigment-10 (peak 10) = unidentified anthocyanin.

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10. All of *Pharbitis nil* the mutants used were from Dr. Nitasaka's collection at Kyusyu University [see: "The Japanese Morning Glory (=Asagao) Home Page"; <http://mg.biology.kyusyu-u.ac.jp/index.html>]. His collection numbers of the mutants used here are: *duskish-1*, 853; *duskish-2*, 854; *dusky*, 856; *dingy*, 857.
11. HPLC was run on Inertsil ODS-2 column (4.6×250 mm for analysis, and 20×250 mm for prep.) at 35°C, and monitoring at 530 nm. Solvent systems for analysis were as follows: linear gradient elution for 40 min from 40 – 85% solvent B (1.5% H<sub>3</sub>PO<sub>4</sub>, 20% AcOH, 25% MeCN) in solvent A (1.5% H<sub>3</sub>PO<sub>4</sub>).
12. Pigment-1 (**1**) = pelargonidin 3-sophoroside-5-glucoside: UV λ<sub>max</sub> (0.1% HCl-MeOH) 507, 268 nm; HPLC Rt (min) 5.1.
13. Pigment-2 (**2**) = peonidin 3-sophoroside-5-glucoside: UV λ<sub>max</sub> (0.1% HCl-MeOH) 522, 277 nm; HPLC Rt (min) 5.5.
14. Pigment-6 (**3**) = pelargonidin 3-*O*-[2-*O*-(β-D-glucopyranosyl)-6-*O*-(*trans*-4-*O*-(β-D-glucopyranosyl)-caffeoyl)-β-D-glucopyranoside]-5-*O*-β-D-glucopyranoside: UV λ<sub>max</sub> (0.1% HCl-MeOH) 510, 322, 281 nm, E<sub>acyl</sub>/E<sub>max</sub> = 0.68; HPLC Rt (min) 11.8. <sup>13</sup>C NMR (125.65 MHz) (CF<sub>3</sub>CO<sub>2</sub>D-DMSO-d<sub>6</sub>,

1:9):  $\delta$  pelargonidin: 165.3 (C-2), 146.8 (C-3), 133.1 (C-4), 111.8 (C-4a), 155.4 (C-5), 105.0 (C-6), 167.5 (C-7), 96.4 (C-8), 162.8 (C-8a), 121.0 (C-1'), 135.2 (C-2'), 117.1 (C-3'), 155.4 (C-4'), 116.3 (C-5'), 128.8 (C-6'); caffeic acid: 125.6 (C-1), 116.7 (C-2), 145.2 (C-3), 147.3 (C-4), 118.6 (C-5), 121.0 (C-6), 121.0 (C- $\alpha$ ), 144.2 (C- $\beta$ ), 166.3 (C=O); Glc A: 99.3 (C-1), 79.5 (C-2), 69.9 (C-4), 62.9 (C-6); Glc B: 103.1 (C-1), 69.9 (C-4), 60.8 (C-6); Glc C: 101.7 (C-1), 69.9 (C-4), 60.8 (C-6); Glc D: 101.7 (C-1), 69.9 (C-4), 60.8 (C-6), 77.5-77.1 (4 $\times$ C), 76.3-75.8 (4 $\times$ C), 74.4 (2 $\times$ C), 73.4 (2 $\times$ C). HRFAB-MS [Calcd for C<sub>48</sub>H<sub>57</sub>O<sub>28</sub> requires: 1081.3036. Found: 1081.3014].

15. Pigment-7 (**4**) = peonidin 3-*O*-[2-*O*-( $\beta$ -D-glucopyranosyl)-6-*O*-( $\beta$ -D-glucopyranosyl)caffeoyl- $\beta$ -D-glucopyranoside]-5-*O*- $\beta$ -D-glucopyranoside (Pharbitis Blue anthocyanin 2): UV  $\lambda_{\max}$  (0.1% HCl-MeOH) 525, 324, 280 nm, Eacyl/E<sub>max</sub> = 0.40; HPLC Rt (min) 12.7.
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20. Pigment-3 = pelargonidin 3-sophoroside: UV  $\lambda_{\max}$  (0.1% HCl-MeOH) 509, 270 nm; HPLC Rt (min) 6.9.
21. Pigment-4 = pelargonidin 3-glucoside: UV  $\lambda_{\max}$  (0.1% HCl-MeOH) 509, 270 nm; HPLC Rt (min) 9.3.
22. Pigment-8 = pelargonidin 3-[tri(glucosylcaffeoyl)sophoroside]-5-glucoside (Pharbitis Red anthocyanin 1): UV  $\lambda_{\max}$  (0.1% HCl-MeOH) 515, 318, 288 nm; Eacyl/E<sub>max</sub> = 1.16; HPLC Rt (min) 16.9.
23. Pigment-9 = pelargonidin 3-[2-glucosylcaffeoylglucosyl-6-(caffeoylglucosylcaffeoyl)glucoside]-5-glucoside (Ipomoea Red anthocyanin 1): UV  $\lambda_{\max}$  (0.1% HCl-MeOH) 515, 318, 288 nm; Eacyl/E<sub>max</sub> = 1.06; HPLC Rt (min) 18.1.